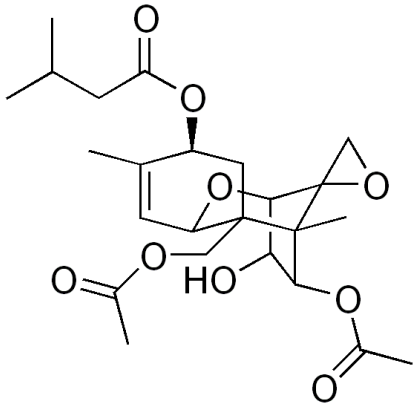
SENSI*Spec* ELISA T-2 Toxin RAPID 96 Tests

Enzyme Immunoassay for the Rapid Quantitative Determination of T-2 Toxin in Food (Cat.nr. HU0030089)

Sensitivity	5 – 16 ppb
Recovery (spiked samples)	92-105%
Incubation Time	20 min

1. GENERAL INFORMATION



T-2 Toxin in addition to deoxynivalenol, zearalenone, the fumonisines and other trichothecenes belongs to the fusarium toxins. These toxins are already produced on the field in consequence of a contact of the cereals by fusarium species. Acute toxic dosages can result in gastroenteritis, damage of bone marrow up to necroses of skin and respiratory passages.

T-2 toxin has a high stability against temperature and can therefore also be detected in bakery products. In Russia the legislator set limit values for T-2 Toxin in food between 50 and 100 ppb. The introduction of limit values in the European union is discussed since many years. Thus an observation of food and feed with respect to the concentration of T-2 Toxin is increasingly obligatory.
The **SENSI*Spec* T-2 Toxin RAPID ELISA** represents a highly sensitive detection system and is particulary capable of the rapid quantification of T-2 Toxin contaminations in cereals, beer, milk and meat.

2. PRINCIPLE OF THE TEST

The **SENSI*Spec* T-2 Toxin RAPID** quantitative test is based on the principle of the enzyme-linked immunosorbent assay. An antibody binding protein is coated on the surface of a microtiter plate. The standards and samples respectively are pipetted together with a T-2 Toxin-peroxidase conjugate and a rabbit-anti-T-2 Toxin antibody into the appropriate wells. The conjugate competes with the T-2 Toxin of samples/standards for the limited number of antibody sites. Simultaneously the anti-T-2 Toxin antibody is bound to the antibody-binding protein coated on the microtiter plate. After 10 min incubation at room temperature, the wells are washed with diluted washing solution to remove unbound material. A substrate solution is added and incubated for 10 minutes, resulting in the development of a blue colour. The colour development is inhibited by the addition of a stop solution, and the colour turns yellow. The yellow colour is measured photometrically at 450 nm. The concentration of T-2 Toxin is indirectly proportional to the colour intensity of the test sample.

3. PRECAUTIONS

Full compliance of the following good laboratory practices (GLP) will determine the reliability of the results:

1) Prior to beginning the assay procedure, bring all reagents to room temperature (20-25°C).

2) All reagents should be mixed by gentle inversion or swirling prior to use. Do not induce foaming.

3) Once the assay has been started, all subsequent steps should be completed without interruption and within the recommended time limits.

4) Replace caps in all the reagents immediately after use. Do not interchange vial stoppers.

5) Use a separate disposable tip for each specimen to prevent cross-contamination.

6) All specimens and standards should be run at the same time, so that all conditions of testing are the same.

7) Do not mix components from different batches.

8) Do not use reagents after expiration date.

9) Check both precision and accuracy of the laboratory equipment used during the procedure (micropipets, ELISA reader etc.).

4. HEALTH AND SAFETY INSTRUCTIONS

1) Do not smoke or eat or drink or pipet by mouth in the laboratory.

2) Wear disposable gloves whenever handling patient specimens.

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3) Avoid contact of substrate and stop solution with skin and mucosa (possible irritation, burn or toxicity hazard). In case of contact, rinse the affected zone with plenty of water.

4) Handling and disposal of chemical products must be done according to good laboratory practices (GLP).

5. REAGENTS

The kit contains reagents for 96 determinations. They have to be stored at 2-8°C. Expiry data are found on the labels of the bottles and the outer package.

1) Microtiter plate consisting of 12 strips with 8 breakable wells each, coated with antibody-binding protein.

2) T-2 Toxin Standards (0; 17.5; 87.5; 350; 875; 1750 ppb): 6 vials with 1 mL each, dyed red, ready-to-use. Because of the total dilution of 1:35 of the solid samples in the extraction step, the calibrators contain 1/35th of the stated value. Thus no further calculation after analysis is necessary.

3) Anti-T-2 Toxin Antibody (rabbit): 6 mL, dyed blue, ready-to-use.

4) Conjugate (T-2 Toxin-Peroxidase): 6 mL, dyed red, ready-to-use.

5) Substrate Solution (TMB): 15 mL, ready-to-use.

6) Stop Solution (0.5 M H_2SO_4): 15 mL, ready-to-use.

7) Sample Diluent (PBS): 2 x 60 mL, dyed red, ready-to-use.

8) Washing Solution (PBS + Tween 20): 60 mL as 10x concentrate. Dilute 1+9 with distilled water. If during the cold storage crystals precipitate, the concentrate should be warmed up to 37°C for 15 minutes.

9) Plastic bag to store unused microtiter strips.

10) Instruction Manual.

6. ADDITIONAL INSTRUMENTATION AND REAGENTS (not provided)

Instrumentation

- 50, 100, 500 and 1000 µL-micropipets
- ELISA reader (450 nm)
- Centrifuge
- Ultra-Turrax, mixer, vortex

Reagents

- Double distilled water
- Methanol

7. SAMPLE PREPARATION

Cereals / Meat

- Grind sample to pass through a 20 mesh sieve and thoughly mix prior to sub-sampling.
- Suspend 20 g of sample in 100 mL of 70% methanol.
- Mix suspension for 5 minutes.
- Filter through Whatman #1 filter or alternatively centrifuge at a minimum of 3000 g for 5 minutes.
- Dilute 100 µL of filtrate/supernatant with 600 µL of sample diluent and test the sample in the ELISA.

Beer / Gyle / Milk

- Dilute an adequate volume of sample diluent with 10% methanol.
- Carbonized beer samples should be preliminarily degassed by moderate heating.
- Cloudy beers (such as beer brewed from wheat) / gyle should preliminarily be sterile-filtered.
- Degrease whole milk samples by centrifugation
- Dilute 100 µL of sample with 900 µL sample diluent/methanol dilution.

  In case of too high concentrated samples, an adequate volume of sample diluent is diluted with 10% methanol. The sample extracts have to be further diluted with this dilution.

8. PROCEDURE

1) Prepare samples as described above.

2) Pipet 100 µL standards or prepared samples in duplicate into the appropriate wells of the microtiter plate. Immediately add 50 µL T-2 Toxin-peroxidase conjugate and 50 µL anti-T-2 Toxin antibody into each well (consider sequence!).

3) Incubate for 10 minutes at room temperature.

4) Wash the plate three times as follows: Discard the contents of the wells (dump or aspirate). Pipet 300 µL of diluted washing solution into each well. After the third repetition empty the wells again and remove residual liquid by striking the plate against a paper towel. The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbencies.

5) Pipet 100 µL of substrate solution into each well.

6) Allow the reaction to develop in the dark (e.g. cupboard or drawer; the chromogen is light-sensitive) for 10 minutes at room temperature.

7) Stop enzyme reaction by adding 100 µL of stop solution (0.5 M H_2SO_4) into each well. The blue colour will turn yellow upon addition.

8) After thorough mixing, measure absorbance at 450 nm (reference wavelength 620 nm), using an ELISA reader. The colour is stable for 30 minutes.

9. CALCULATION OF RESULTS

The ready-to-use standards are prepared for a direct determination of the sample concentrations. The dilution of samples in the extraction process as described in the above stated sample preparation procedure is already considered. Additional dilution due to high sample concentration has to be accounted for.

1) Calculate the average optical density (OD 450 nm) for each set of reference standards or samples.

2) Construct a standard curve by plotting the mean optical density obtained for each reference standard against its concentration in ppm on semi-log graph paper with the optical density on the vertical (y) axis and the concentration on the horizontal (x) axis. Alternatively the evaluation can be carried out by software. In this case the 4-parameter method should be preferred.

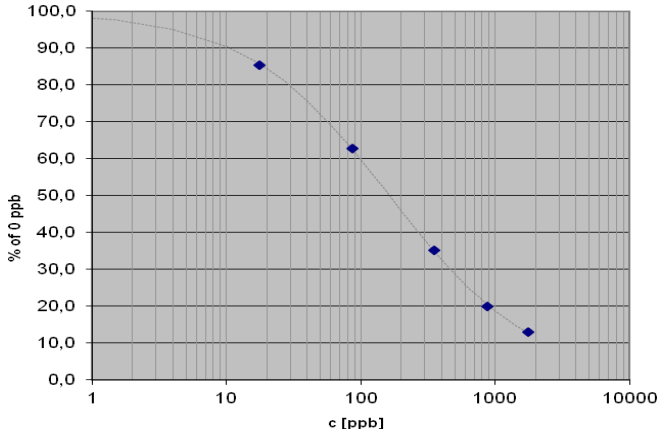
3) Using the mean optical density (OD) value for each sample, determine the corresponding concentration of fumonisin in ppm from the standard curve. Depending on experience and/or the availability of computer capability, other methods of data reduction may be employed.

4) Due to a deviating sample preparation process the results for Beer / Gyle / Milk samples additionally have to be multiplied with 0.286 in order to get the real concentration of the sample.

10. TYPICAL STANDARD VALUES

The following table contains an example for a typical standard curve. The binding is calculated as percent of the absorption of the 0 ppb standard. These values are only an example and should not be used instead of the standard curve which has to be measured in each new test.

T-2 Toxin (ppb)	(% binding of 0 ppb)
0	100
17.5	85
87.5	62
350	35
875	20
1750	13



11. PERFORMANCE

Sensitivity

The limit of detection (LOD) of the **SENSI*Spec* T-2 Toxin RAPID test** is 13 ppb.

Validation experiments with common matrices resulted in the following LODs [ppb].

Wheat	10
Rye	11
Barley	16
Corn	14
Rice	11
Meat (pork)	8
Milk	5
Beer	5

The limit of quantification (LOQ) of the **SENSI*Spec* T-2 Toxin RAPID test** is 17.5 ppb.

Due to the variety of sample matrices and their influence on the blank, results less than the LOQ should be treated as negative.

11.1. Cross-reactivity

Cross-reactivity relative to T-2 Toxin(=100%)

HT-2 Toxin	3.0%
T-2 Triol	0.35%
T-2 Tetraol	0.07%

11.2. Precision

Intra-assay Precision	3-4%
Inter-assay Precision	3-6%

11.3. Linearity

The serial dilution of spiked samples (wheat, rice, corn, rye, barley, meat, beer, milk) resulted in a dilution linearity of 84-111%.

11.4. Recovery

Wheat	100%
Barley	96%
Rye	103%
Oats	97%
Rice	95%
Corn	92%
Meat	97%
Beer	105%
Milk	95%

12. REFERENCES

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