

Celer AFLA M₁ 500

Enzyme immunoassay for the detection of Aflatoxin M₁ (Cat.nr. HU0040081)

Celer AFLA M₁ 500 is a kit prepared for a competitive immunoenzymatic assay for the quantitative analysis of aflatoxin M₁.

The kit contains the procedure and the materials sufficient for 96 determinations including standards. For result evaluation a microtiter plate or a strip photometer is required (manual or automatic ELISA reader).

Type of samples that can be analyzed (matrices)

Raw milk, milk powder.

Sample preparation

- Raw milk: no preparation required.
- Milk powder: dilution.

Assay time: 20 minutes (sample preparation not included).

Detection limit

- Raw milk: 25 ng/l
- Milk powder: 250 ng/l

Specificity

Compound	Cross-reactivity (%)
Aflatoxin M ₁	100

1. TEST PRINCIPLE

The assay is performed in plastic microwells which have been coated with anti-Aflatoxin M₁ antibodies. Aflatoxin M₁ standard solutions and samples are added to the microwells.

During the first incubation, free Aflatoxin M₁ molecules are bound to the anti-Aflatoxin M₁ antibodies.

Any unbound substance is then removed in a washing step.

A second incubation is performed with an aflatoxin-HRP conjugate, which covers all the remaining free binding sites of the antibody. After the incubation a second washing step is performed.

The bound enzyme activity is determined by adding a fixed amount of a chromogenic substrate. The enzyme converts the colorless chromogen into a blue product during the third incubation.

The addition of the stop reagent leads to a color change from blue to yellow. The absorbance is measured by a microplate reader at 450 nm. The color development is inversely proportional to the Aflatoxin M₁ concentration in the sample.

2. PROVIDED REAGENTS

Microtiter plate: 96 wells (12 strips x 8 wells) coated with anti-Aflatoxin M₁ antibodies.

As the strips are breakable, the wells can be used individually. For this purpose, it is sufficient to take out the wells from the frame and to break the joint.

Aflatoxin M₁ standard solutions: 5 plastic vials containing 1.5 ml of: 0 ng/l; 25 ng/l; 50 ng/l; 200 ng/l; 500 ng/l.

Enzyme conjugate: 1 plastic bottle containing 14 ml.

Washing-buffer 20X: 1 plastic bottle containing 50 ml.

Developing solution: 1 plastic bottle containing 14 ml.

Stop solution: 1 glass bottle containing 8. White cap.

3. MATERIALS REQUIRED BUT NOT PROVIDED

- "Milk Diluent" (cat.nr. HU0040101)
- Distilled water

Equipment

- Balance
- Microplate reader, filter 450 nm
- Micropipette 50-200 µl, tips
- Multichannel micropipette 50-250 µl, if using more than three strips

4. WARNING AND PRECAUTIONS FOR THE USERS

- The test is for *in vitro* diagnostic use only.
- Some reagents contain solutions that may be identified as dangerous substance by the Regulation (EC) N° 1272/2008. Please refer to *Material Safety Data Sheet* available on both the Eurofins Technologies and Eurofins Tecna (tecna.eurofins-technologies.com) web site.
- Handle the reagents with caution, avoiding contact with skin, eyes and mucous membranes.

5. HANDLING AND STORAGE INSTRUCTIONS

- Store the kit at +2/+8 °C and never freeze.
- **Bring all reagents to room temperature before use (2 hours). ATTENTION: Do not unseal the microplate until it reaches the room temperature.**
- Reseal the unused strips of the microtiter plate in the bag together with the desiccant bag provided.
- Return all reagents to +2/+8 °C immediately after use.
- Do not use components after the expiration date.
- Do not intermix components from different kit lots.
- Do not use photocopies of the instruction booklet. Follow the original instruction booklet that is included with the kit.
- Do not change the assay procedure, in particular:
 - do not prolong the incubation times;
 - do not incubate the plate at temperatures higher than 25°C;
 - do not shake the plate during the incubations.
- Use accurate and precise micropipettes with suitable tips for dispensing.
- Once started, complete all the steps without interruption.
- The reproducibility of ELISA results depends largely upon the efficiency and uniformity of microwells washing; always keep to the described procedure.
- Use a single disposable tip for each standard solution and sample to avoid cross-contamination.
- Do not allow tips to contact the liquid already in the microwells.
- Avoid exposure to direct light during all incubations. It is recommended to cover the microtiter plate without using plate sealers.

6. SAMPLES PREPARATION

6.1. Raw milk

After milking, the milk has to be tested within 24 hours. Otherwise milk has to be stabilized (with sodium azide, azidiol or similar substances)

- 1) Bring the milk to room temperature.
- 2) Analyse the milk directly in the assay, without skimming.
- 3) If the sample is dosed >500 ng/l, it is suggested to dilute it four times (1+3) in "Milk Diluent" (cat.nr. HU0040101) and retest it, in order to obtain a dosage range 100-2000 ng/l.

6.2. Powdered milk

- 1) Weight out 10 g of the powder and get to a volume of 100 ml with distilled water (it is suggested to heat the water to 37°/40 °C to dissolve the powder easier).
- 2) Shake until the powder is completely dissolved.
- 3) The dilution factor is 10.

7. WORKING SOLUTIONS PREPARATION

Aflatoxin M1 standard solutions: ready to use (**shake gently prior to use**).

Enzyme conjugate: ready to use.

Washing buffer: dilute the concentrate 1:20 (1+19) with distilled water. **ATTENTION**: If crystals are present, bring the solution to room temperature and stir in order to solve them completely.

The diluted washing buffer is stable at room temperature for 24 hours and at +2/+8°C for two weeks.

Developing solution: ready to use. The solution is light sensitive and must be stored away from direct light.

Stop solution: ready to use. **ATTENTION**: It contains 1M sulphuric acid. Handle with care; in case of contact flush immediately with plenty of water.

8. ASSAY PROCEDURE

- 1) Predispose an assay layout, recording standard solutions and samples positions, taking into account that one well is required for each standard and sample.

ATTENTION: It is suggested to carry out no more than 24 determinations in each assay (standards included).
- 2) First incubation
 - Add 100 µl of each standard/ sample into the corresponding wells.
 - Shake the plate gently with rotatory motion for few seconds
 - Incubate 10 minutes at room temperature.
 - Do not prolong the first incubation time and do not use automatic shakers.
- 3) Washing
 - Pour the liquid out from the wells.
 - Fill completely all the wells with working wash solution using a squeeze bottle. Pour the liquid out from the wells.
 - Repeat the washing sequence three (3) times.
 - Remove the remaining droplets by tapping the microplate upside down vigorously against absorbent paper.

Do not allow the wells to dry out.

- 4) Second incubation
 - Using a multichannel pipet, add to the wells 100 µl of the enzyme conjugate solution.
 - Shake the plate gently with rotatory motion for few seconds.
 - Incubate for 5 minutes.
- 5) Repeat step 3.
- 6) Developing
 - Using the multichannel micropipette, add 100 µl of developing solution to each well.
 - Mix thoroughly with rotatory motion for few seconds.
 - Incubate for 5 minutes at room temperature.
- 7) Using a multichannel pipet, add 50 µl of stop solution to each well and mix thoroughly with rotatory motion for few seconds.
- 8) Measure the absorbance at 450 nm.
- 9) Read within 15 minutes.

9. CALCULATION OF RESULTS

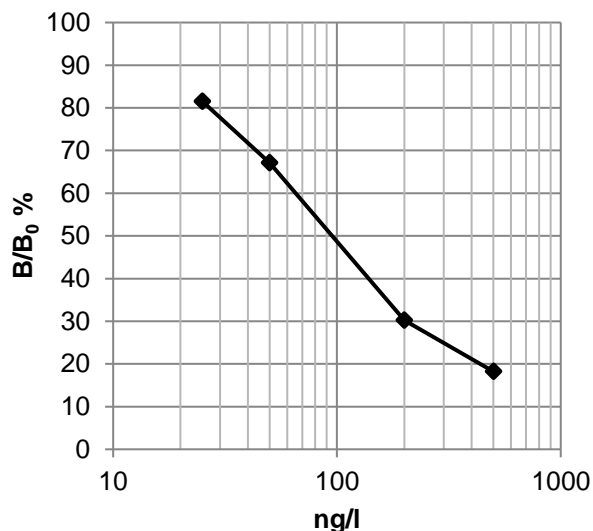
- Calculate the mean absorbance of each, standard and sample.
- Divide the mean absorbance value of each standard and sample by the mean absorbance of the standard 0 and multiply by 100; the standard 0 is thus made equal to 100% and all the other absorbance values are expressed as percentages:

$$\frac{\text{Standard (or sample) absorbance}}{\text{Standard 0 (B}_0\text{) absorbance}} \times 100 = \frac{B}{B_0} (\%)$$

- Enter the B/ B₀ values calculated for each standard in a semi-logarithmic system of coordinates and draw the standard curve.
- Interpolate the B/ B₀ value of each sample to the corresponding concentration from the calibration curve. For dilution applications multiply this concentration for the dilution factor.

Please note: to build the calibration curve, the “spline” algorithm can be used, but it is also possible to use the “point to point” curve. To elaborate the ELISA results using the “point to point” method, Excel spreadsheet are available on Eurofins Tecna website tecna.eurofins-technologies.com and can be downloaded directly from the bottom of the product page.

10. CALIBRATION CURVE EXAMPLE



11. EVALUATION OF RESULTS

After processing the results, it is necessary to verify the assay performance. The verification is performed by comparison of obtained data with those given in kit specifications (see chapter 12).

If the values are outside the specifications given, then the results of the test are not assured, therefore the aflatoxin M₁ concentration levels in the samples may not be valid.

In these cases it is advised to check the expiry date of the kit, the wavelength at which the reading was performed, as well as the procedure followed.

If operation errors are not identified as cause, contact our technical assistance.

WARNING: Kit replacement will only be possible in case of return. The kit must be stored in its integral version at +2/+8°C.

12. KIT SPECIFICATIONS

12.1. Assay specification

Description	Specifications
Mean B ₀ absorbance	≥ 0.7 OD _{450nm}
B/ B ₀ 50 %	54-110 ng/l

13. LIABILITY

Samples evaluated as positive using the kit have to be re-tested with a confirmation method.

Eurofins Technologies Hungary shall not be liable for any damages to the customer caused by the improper use of the kit and for any action undertaken as a consequence of results.

Eurofins Technologies Hungary shall not be liable for the unsafe use of the kit out of the current European safety regulations.